# **CASE REPORT**



# A case report: Marfan syndrome with X trisomy and FBN1 and SDHB mutations

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# Abstract

**Background** Marfan syndrome (MFS) is a rare autosomal dominant connective tissue disorder affecting the cardiovascular, skeletal, and ophthalmic systems. This report aimed to describe a novel genetic background and treatment prognosis of MFS.

**Case presentation** A proband was initially diagnosed with bilateral pathologic myopia and suspected MFS. We performed whole exome sequencing and found a pathogenic nonsense *FBN1* mutation in the proband, which confirmed the diagnosis of MFS. Notably, we identified a second pathogenic nonsense mutation in *SDHB*, which increased the risk of tumours. In addition, the proband karyotype was X trisomy, which may cause X trisomy syndrome. At the 6-month follow-up after posterior scleral reinforcement surgery, the proband's visual acuity improved significantly; however, myopia was still progressing.

**Conclusions** We report a rare case of MFS with a X trisomy genotype, a mutation in *FBN1* and a mutation in *SDHB* for the first time, and our findings could be helpful for the clinical diagnosis and treatment of this disease.

Keywords Marfan syndrome, X trisomy, FBN1, SDHB, Case report

# Background

MFS is a rare autosomal dominant connective tissue disease with an incidence of approximately 1 in 5000, and its clinical symptoms mainly involve the cardiovascular, skeletal, and ophthalmic systems [1]. In most cases, MFS is caused by mutations in *FBN1*. *FBN1* encodes the major component of extracellular microfibrils, which interfere with local transforming growth factor  $\beta$  signalling and disrupt tissue integrity [2]. Moreover, gene mutation

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<sup>3</sup> Quanzhou Women's and Children's Hospital, Quanzhou, People's Republic of China detection is suitable for early diagnosis if clinical symptoms do not meet diagnostic criteria [3]. Rare genetic events and novel mutations that are disease associated (particularly for monogenic diseases) and potential disease-causing mutations due to incomplete penetrance can be detected by whole exome sequencing (WES) [4]. In this study, we aimed to describe the specific genetic background and treatment prognosis of MFS.

# **Case presentation**

A 4-year-old girl with normal weight (18 kg) who was slightly taller than girls of the same age (118 cm) complained at the initial consultation. Poor binocular vision was found during physical examination 1 year prior, with no other abnormalities detected. The patient's myopia gradually progressed, and she came to our hospital for treatment. The preliminary diagnosis was bilateral pathologic myopia and suspected MFS. Past history revealed no hypertension, diabetes, other eye diseases, infectious diseases, trauma, or surgery. Birth history: the patient



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was born via a full-term natural delivery at a birth weight of 2.8 kg, with no history of asphyxia after birth. The G1P1 mother was healthy during pregnancy and denied any history of medication use and X-ray exposure.

Preoperative physical examination: The patient exhibited stable vital signs. The visual acuity with glasses was 0.25 (mydriatic refraction,  $-7.00/-1.50*15 \rightarrow 0.2$ ) for oculus dexter (OD) and 0.25 (mydriatic refraction,  $-4.00/-0.75*160 \rightarrow 0.25$ ) for oculus sinister (OS). The intraocular pressure was 11 mmHg for OD and 13 mmHg for OS. The axial length was 25.49 mm in the right eye and 23.95 mm in the left eye. Other ophthalmic examinations, including lentis, were normal. Auxiliary examination: Routine electrocardiogram showed sinus bradycardia with arrhythmia, as shown in Fig. 1. Echocardiography revealed that because the diameter of the aortic Valsalva sinus was 2.18 cm, which is greater than the upper limit of the reference value of 2.0 cm, the aortic sinus was slightly dilated, as shown in Fig. 2. The aortic root Z value of 2.078 was slightly greater than the upper limit of the reference value of 2, which suggested aortic dilatation, and left ventricular systolic function was normal. Routine haematuria, coagulation, biochemical, and chest X-ray evaluations revealed no abnormalities.

After hospitalization, we performed posterior scleral reinforcement surgery, which was successful. Condition at discharge: visual acuity, OD: 0.4 and OS: 0.5; intraocular pressure, 15 mmHg OD and 15 mmHg OS; mild oedema of the eyelids; and hyperaemia and oedema of the conjunctiva (+). Other eye examinations were



Fig. 2 Echocardiographic image of aortic root. *Note*: the red arrow indicates aortic sinus of Valsalva

normal. To confirm the diagnosis, peripheral blood was drawn from the proband for WES.

At follow-up after 6 months, the proband's visual acuity with glasses was 0.5 (mydriatic refraction, -8.50/-2.50\*15) for OD and 0.6 (mydriatic refraction, -4.50/-1.00\*155) for OS. The axial length was 25.98 mm in the right eye and 24.28 mm in the left eye. Her parents were basically satisfied with the results of the operation.

## **Genetic analysis**

WES was performed using the Illumina NovaSeq PE150 platform. Sequencing quality control parameters were as follows: the coverage of the target region was>99.69%, and the average sequencing depth of the target region was 144.14×. We sequenced the entire human exome,



Fig. 1 Detection record of routine electrocardiogram

Genetic pattern	AD	AD
Related diseases (OMIM)	MFS (154,700)	Paraganglioma type 4 (115,310)
ACMG Pathogenicity RATING	Pathogenic	Pathogenic
Mother	Wild type	Wild type
Father	Wild type	Heterozygous
Proband	Heterozygous	Heterozygous
Types of mutation	Nonsense	Nonsense
Variant site	NM_000138: exon65: c.8080C > T: p.Arg2694*	NM_003000: exon2: c.136C >T: p.Arg46*
rs number	rs 200309328	rs 74315370
Chromosomal location	chr15: 48704912	chr1: 17371320
Gene	FBN1	SDHB

 Table 1
 Information on gene variant sites

ACMG American College of Medical Genetics and Genomics, AD autosomal dominant

\* indicates that an amino acid at a certain position becomes a stop codon

covering>20,000 genes and>85% of human genetic diseases. Read-depth-based copy number variation (CNV) was analysed by CNVkit software (https://cnvkit.readt hedocs.io/en/stable/). A copy number of 3 was considered a triploid duplicate.

The proband phenotype (high myopia, sinus bradycardia with arrhythmia, slightly dilated aortic sinus, slender lower extremities, and loose ligaments of the limbs) was consistent with MFS caused by an FBN1 mutation. In particular, we identified a nonsense mutation in exon 65 of FBN1 caused by the substitution of cytosine (C) to thymine (T) at position 8080 of the cDNA. This mutation resulted in the substitution of arginine (Arg) to a stop codon at position 2694, as shown in Table 1. As demonstrated in Fig. 3, Sanger sequencing was used to confirm the FBN1 mutation. Moreover, pedigree analysis revealed that this heterozygous variant in the proband was a de novo variant. According to the 2015 American College of Medical Genetics and Genomics (ACMG) guidelines, the pathogenic evidence of this variant was "PVS1+PS2+PS4+PM2\_Supporting;" thus, we classified it as pathogenic. Additionally, the heterozygous *FBN1* variant in the proband conformed to the autosomal dominant (AD) inheritance pattern of MFS. The nonsense FBN1 mutation confirmed the diagnosis of MFS.

In addition to the FBN1 mutation, we identified two other genetic alterations. The first was an single nucleotide variation in SDHB, which was a secondary finding because the clinical phenotype of the SDHB mutation did not correspond to the clinical symptoms of the proband. In particular, we identified a nonsense mutation in exon 2 of SDHB caused by the substitution of C to T at position 136 of the cDNA, resulting in the substitution of Arg at position 46 to a stop codon, as shown in Table 1. As demonstrated in Fig. 4, Sanger sequencing was used to confirm the SDHB mutation. Pedigree analysis revealed that this heterozygous mutation was inherited from the father and that it conformed to the AD inheritance pattern of paraganglioma (PGL) type 4 and pheochromocytoma (PCC). Therefore, the variant was classified as pathogenic (PVS1+PS4+PM2\_Supporting), according to the ACMG guidelines. The other genetic alteration we identified was a CNV, particularly a copy number duplication at the Xp22.33-Xq28 region containing 1154 genes, as shown in Table 2 and Fig. 5. This 155 Mb duplication was classified as pathogenic according to the 2019 ACMG criteria for CNVs. The genetic syndrome associated with this region is Superfemale syndrome.

# **Discussion and conclusions**

MFS involves multiple systems [5], and vision loss and skeletal system abnormalities are typically detected earlier than cardiovascular abnormalities [4]. These features are consistent with this case, as severe vision loss was the first symptom we detected in this proband. A previous study revealed that posterior scleral reinforcement surgery in patients with MFS and retinoschisis results in retinal reattachment [6]. Notably, the proband's visual acuity improved significantly after surgery; in particular, it increased from 0.25 to 0.5 OD and from 0.25 to 0.6 OS. However, myopia progressed from -7.00 to -8.50OD (mydriatic refraction) and from -4.00 to -4.50 OS (mydriatic refraction). One possible explanation is that surgery promoted retinal angiogenesis but could not inhibit axial growth, as the axial length increased from 25.49 to 25.98 mm in the right eye and from 23.95 to 24.28 mm in the left eye.

According to the Online Mendelian Inheritance in Man (OMIM) database, FBN1 mutations (OMIM\*134,797) can cause eight diseases, including acromicric dysplasia, MFS, and stiff skin syndrome. We diagnosed MFS based on the proband phenotype, which was consistent with that of MFS. In addition to being associated with FBN1, MFS is also associated with mutations in TGFBR1 and TGFBR2. Through WES technology, more genes that may be related to MFS, such as TTN and POMT1, can be identified [7], and a small, 0.76 Mb microdeletion of CNV in 15q21.1 leading to haploinsufficiency of the fibrillin 1 (FBN1) gene associated with MFS was discovered [8]. In view of the advantages of WES in the diagnosis of MFS, WES was used for the genetic diagnosis of this proband. The heterozygous FBN1 variant p.R2694\* we identified in the proband has a 50% chance of being inherited by the next generation. Since the FBN1 mutation was a de novo variant, the likelihood of this mutation occurring in the parents' next child is very small, considering the possibility of germ cell mosaicism in one parent.

SDHB mutations (OMIM\*185,470) are related to gastrointestinal stromal tumours, mitochondrial complex II deficiency, PGL, gastric stromal sarcoma, and PCC and are associated with a high rate of malignancy in PGL and PCC [9]. The overall penetrance of SDHB mutations is approximately 21% at the age of 50 and 42% at the age of 70 [10]. This is likely why the proband and her father had no symptoms of PGL and PCC. We recommend follow-up of the proband and her father to assess the clinical phenotype and perform relevant examinations, if necessary, with the aim of further clarifying the clinical significance of SDHB mutations. X trisomy syndrome was first described by Jacobs et al. [11], According to recent statistics, the incidence of X trisomy syndrome is approximately 11 in 10,000 women, and only approximately 13% of patients are diagnosed [12]. Females with X trisomy syndrome have a relatively mild form of the disease and do not have typical phenotypic features, which could explain why the diagnosis

Sample	Genotype	<i>FBN1</i> : NM_000138: c.8080C>T: p.Arg2694X
Proband	Heterozygosity	NCBI GRCh37 reference: G G G C A T G G G C C G A G G A A C C C Sangar sequence: G G G C A T G G G C T G A G G A A C C C
		AAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAA
Father	Wild type	NCBI GRCh37 reference: G G G T T T C C T C G G C C C A T G C C C Sunger sequence: G G G T T T C C T C G G C C C A T G C C C A
Mother	Wild type	NCBI GRCh37 reference: G G G C A T G G G C C G A G G A A A C C C Sanger sequence: G G G C A T G G G C C G A G G A A A C C C

Fig. 3 Validation of FBN1 gene mutation

Sample	Genotype	<i>SDHB</i> : NM_003000: c.136C>T: p.Arg46X
Proband	Heterozygosity	NCBI GRCh37 reference: GGGTCCCATCGATAGATGGCA Sanger sequence: GGGTCCCATCAATAGATGGCA
Father	Heterozygosity	NCBI GRCh37 reference: GGGTCCCATCGATAGATGGCA Senger sequence: GGGTCCCATCAATAGATGGCA
Mother	Wild type	NCBI GRCh37 reference: G G G T C C C A T C G A T A G A T G G C A Sanger sequence: G G G T C C C A T C G A T A G A T G G C A MMMMMMMMMMMMMMMMMMMMMMMMMMMMMMMMMMMM

Fig. 4 Validation of SDHB gene mutation

Table 2 D	Details of the	repeating re	aion of tl	he break	point	of the X	chromosome
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Chromosomal region	CNV type	Fragment size (bp)	Chromosomal location	Contains major genes	
Xp22.33-Xq28	Duplication (CNV = 3)	155,072,003	chrX: 60,500 -155,132,503	ABCB7, ABCD1, ACE2, ACOT9, ACSL4, ACTRT1, ADGRG2	
				ADGRG4,AFF2,AGTR2 etc. including 1154 genes	

Xp22.33-Xq28 covers the entire X chromosome



Fig. 5 Detection results for 3 chromosome X. *Note*: the abscissa represents the schematic diagram of the position of the chromosome X. Yellow dots represent detection sites. The copy number 3 on the vertical axis means that the number of chromosome X is 3

is easily missed [13]. This 4-year-old proband did not show X trisomy syndrome. In contrast to the symptoms reported in the two previous cases of MFS with X trisomy syndrome [14, 15], we found no immune abnormalities or dysmorphia in the proband, likely due to different genetic alterations. The limitation of this study is that trisomy X was not further verified by karyotype analysis.

In conclusion, this proband was diagnosed with MFS with the X trisomy genotype, a mutation in the *FBN1* gene and a mutation in *SDHB*. Our findings could be helpful for the clinical diagnosis and treatment of this disease.

### Abbreviations

MFS	Marfan syndrome
WES	Whole exome sequencing
OD	Oculus dexter
OS	Oculus sinister
ACMG	American College of Medical Genetics and Genomics
С	Cytosine
AD	Autosomal dominant
Т	Thymine
Arg	Arginine
PGL	Paraganglioma
PCC	Pheochromocytoma
CNV	Copy number variation

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### Author contributions

JL designed the study and wrote the article. YL designed the study, collected the clinical data and assisted with specimen collection. GW revised and polished the manuscript. All authors approved the final article.

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### Availability of data and materials

The datasets generated during the current study are available in the National Center of Biotechnology Information (NCBI), and the accession number SUB12917819 is under review.

### Declarations

### Ethics approval and consent to participate

This study followed the principles of the Declaration of Helsinki and was approved by the Ethics Committee of the Quanzhou Women's and Children's Hospital. The participant' legal guardian provided signed written informed consent to participate in this study.

### **Consent for publication**

Written informed consent was obtained from the participants' legal guardian for publication of this case report and any accompanying details and images. A copy of the written consent is available for review by the Editor of this journal.

### **Competing interests**

The authors declare no conflict of interest.

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### References

- 1. Judge DPDH. Marfan's syndrome. Lancet. 2005;366(9501):1965-76.
- Coelho SGAA. Marfan syndrome revisited: from genetics to the clinic. Rev Port (Cardiol Engl Ed). 2020;39(4):215–26.
- 3. Sheikhzadeh SKC, Keyser B, Stuhrmann M, Arslan-Kirchner M, Rybczynski M, Bernhardt AM, Habermann CR, Hillebrand M, Mir T, Robinson PN,

Berger J, Detter C, Blankenberg S, Schmidtke J, von Kodolitsch Y. Analysis of phenotype and genotype information for the diagnosis of Marfan syndrome. Clin Genet. 2012;82(3):240–7.

- 4. Du QZD, Zhuang Y, Xia Q, Wen T, Jia H. The molecular genetics of Marfan syndrome. Int J Med Sci. 2021;18(13):2752–66.
- Zeigler SM, Sloan B, Jones JA. The pathophysiology and pathogenesis of Marfan syndrome. Adv Exp Med Biol. 2021;1348:185–206.
- He QWR. Comparison of biometric and structural ocular manifestations of Marfan syndrome in twins. Zhonghua Yan Ke Za Zhi. 2021;57(7):540–3.
- Lin MRCC, Ting J, Chang JG, Chou WH, Huang KJ, Cheng G, Chang HH, Chang WC. Application of whole exome sequencing and functional annotations to identify genetic variants associated with Marfan syndrome. J Pers Med. 2022;12(2):198.
- Yang R, Zhang W, Lu H, Liu J, Xia Y, Liao S, Li X, Zhang X, Fan X, Wang C. Identification of a novel 15q21.1 microdeletion in a family with Marfan syndrome. Genet Res (Camb). 2022;2022:3556302.
- 9. Bholah RBT. Review of pediatric pheochromocytoma and paraganglioma. Front Pediatr. 2017;5:155.
- Rijken JANNJM, Eijkelenkamp K, Jansen JC, van Berkel A, Timmers HJLM, Kunst HPM, Bisschop PHLT, Kerstens MN, Dreijerink KMA, van Dooren MF, van der Horst-Schrivers ANA, Hes FJ, Leemans CR, Corssmit EPM, Hensen EF. The penetrance of paraganglioma and pheochromocytoma in SDHB germline mutation carriers. Clin Genet. 2018;93(1):60–6.
- Jacobs P, Baikie AG, Brown WC, Macgregor TN, Maclean N, Harnden DG. Evidence for the existence of the human "super female." Lancet. 1959;2(7100):423–5.
- Berglund AVM, Skakkebæk A, Chang S, Stochholm K, Gravholt CH. Changes in the cohort composition of turner syndrome and severe non-diagnosis of Klinefelter, 47, XXX and 47, XYY syndrome: a nationwide cohort study. Orphanet J Rare Dis. 2019;14(1):16.
- Manning M, Hudgins L. Array-based technology and recommendations for utilization in medical genetics practice for detection of chromosomal abnormalities. Genet Med. 2010;12(11):742–5.
- 14. Smith TFEE. Marfan's syndrome with 47, XXX genotype and possible immunologic abnormality. South Med J. 1981;74(5):630–2.
- Tinsa FKL, Chekib J, Hamouda S, Ben Jemaa L, Boussetta K, Bousnina S. Marfan syndrome in a Triple-X girl: a new association? Tunis Med. 2010;88(3):203–6.

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