

Blood-based DNA methylation and exposure risk scores predict PTSD with high accuracy in military and civilian cohorts

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Abstract

Background Incorporating genomic data into risk prediction has become an increasingly popular approach for rapid identifcation of individuals most at risk for complex disorders such as PTSD. Our goal was to develop and validate Methylation Risk Scores (MRS) using machine learning to distinguish individuals who have PTSD from those who do not.

Methods Elastic Net was used to develop three risk score models using a discovery dataset (n=1226; 314 cases, 912 controls) comprised of 5 diverse cohorts with available blood-derived DNA methylation (DNAm) measured on the Illumina Epic BeadChip. The frst risk score, exposure and methylation risk score (eMRS) used cumulative and childhood trauma exposure and DNAm variables; the second, methylation-only risk score (MoRS) was based solely on DNAm data; the third, methylation-only risk scores with adjusted exposure variables (MoRSAE) utilized DNAm data adjusted for the two exposure variables. The potential of these risk scores to predict future PTSD based on pre-deployment data was also assessed. External validation of risk scores was conducted in four independent cohorts.

Results The eMRS model showed the highest accuracy (92%), precision (91%), recall (87%), and f1-score (89%) in classifying PTSD using 3730 features. While still highly accurate, the MoRS (accuracy=89%) using 3728 features and MoR-SAE (accuracy=84%) using 4150 features showed a decline in classifcation power. eMRS signifcantly predicted PTSD in one of the four independent cohorts, the BEAR cohort (beta = 0.6839 , p=0.006), but not in the remaining three cohorts. Pre-deployment risk scores from all models (eMRS, beta=1.92; MoRS, beta=1.99 and MoRSAE, beta=1.77) displayed a significant (p < 0.001) predictive power for post-deployment PTSD.

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Conclusion The inclusion of exposure variables adds to the predictive power of MRS. Classifcation-based MRS may be useful in predicting risk of future PTSD in populations with anticipated trauma exposure. As more data become available, including additional molecular, environmental, and psychosocial factors in these scores may enhance their accuracy in predicting PTSD and, relatedly, improve their performance in independent cohorts.

Keywords DNA methylation, Machine learning, PTSD, Risk scores

Background

Posttraumatic stress disorder (PTSD) is a psychiatric disorder that can develop after experiencing or witnessing a life-threatening event such as a war/combat, natural disaster, violence, or serious accident. PTSD occurs in~13% of the trauma-exposed population [[1\]](#page-12-0), and females are twice as likely to experience PTSD as males [[2\]](#page-12-1). PTSD commonly occurs together with other psychiatric disorders [[3–](#page-12-2)[6](#page-12-3)] and has also been associated with other health conditions such as accelerated aging [\[7](#page-12-4), [8](#page-12-5)], cardiovascular and metabolic disorders [\[9](#page-12-6), [10](#page-12-7)], and poor physical health [[11\]](#page-12-8). Consequently, the overall burden caused by PTSD is high, with an estimated annual economic burden of \$232 billion in the United States in 2018, including \$76.1 billion in excess direct health care costs [\[12](#page-12-9)]. Identifying individuals at elevated risk of PTSD would enhance the ability to develop timely preventive strategies and therapies for this disorder.

Incorporating genomic data into risk prediction has become an increasingly popular approach for rapid identifcation of individuals most at risk for complex disorders such as PTSD. In particular, polygenic risk scores (PRS) have been evaluated in both research and clinical contexts to estimate risk to develop complex disorders, including coronary artery disease, breast cancer, Type 2 diabetes, and Alzheimer's Disease (reviewed in [\[13\]](#page-12-10)). These genetically-based risk scores are attractive as they access lifetime risk for a particular disorder and leverage variation across hundreds to thousands of variants. However, most PRSs are not yet clinically useful, as they typically explain only a small proportion of variance in risk for a particular disorder and do not capture environmental factors that infuence risk or detect the efect of disease progression itself [\[14\]](#page-12-11), both of which may be important to identifying individuals at highest risk for disease.

In contrast, risk scores based on DNA methylation (DNAm) levels, which are modifable and dynamic, can potentially convey more information about disease risk. A growing literature has shown that approaches originally developed for generating PRS can be adapted for DNAm data (reviewed in $[15, 16]$ $[15, 16]$ $[15, 16]$ $[15, 16]$ $[15, 16]$). The resulting methylation risk scores (MRS) have been shown in some cases to be more indicative of current disease state [\[17](#page-12-14)] and health-related phenotypes [[18\]](#page-12-15), as well as more predictive of future disease risk [\[19](#page-12-16)], than PRS-based approaches. Indeed, for PTSD, which requires an environmental exposure—trauma/shocking event —to meet the requirements for a diagnosis, MRS-based risk scores that capture the diferential efects of this exposure may be particularly informative for identifying trauma-exposed individuals most at risk for the disorder.

To this end, here we leverage a large, ancestrally diverse set of cohorts to take a frst step toward developing MRS for PTSD. We focus specifcally on developing scores that distinguish between those with vs. without the disorder (i.e., a diagnostic MRS that correctly classifes current cases vs. trauma-exposed controls), and attempt to replicate these MRS in multiple external validation cohorts. We further test whether these diagnostic risk scores have prognostic value, i.e., can predict future PTSD among individuals prior to trauma exposure. Finally, to gain insight into potential mechanisms, we investigate the biological signifcance associated with the specifc cytosineguanine sites separated by a phosphate group (i.e. (CpG) sites) that comprise the MRS.

Methods

Cohorts

In order to maximize the available data from which to develop risk scores using machine learning approaches, we created a discovery cohort comprised of 1226 individuals drawn from fve cohorts (Table [1](#page-2-0)). Two of these cohorts are civilian— Detroit Neighborhood Health Study (DNHS) and Grady Trauma Project (GTP), and three cohorts are military— Army Study to Assess Risk and Resilience in Servicemembers (Army STARRS), Marine Resilience Study (MRS I&II), and Prospective Research in Stress-related Military Operations (PRISMO). Details about each cohort are given in the supplementary file. The overall workflow of the preprocessing and methods combining data from the fve cohorts is shown in supplementary fle (Figure S1).

Quality Control (QC) procedures

DNAm from whole blood was measured using the Illumina MethylationEPIC BeadChip following the manufacturer's recommended protocol. Raw DNAm β values were obtained, and a sex check was conducted using the

	Current PTSD					
	Cases	Controls	P value	Total		
N						
Army STARRS	42	111		153		
DNHS	31	385		416		
GTP	161	323		484		
MRS I&II	63	60		123		
PRISMO	17	33		50		
All	314	912		1226		
Gender, Male (%)						
Army STARRS	42 (27)	111(73)		153 (100)		
DNHS	10(2)	161 (39)		171 (41)		
GTP	25(5)	107(22)		132 (27)		
MRS I&II	63 (51)	60 (49)		123 (100)		
PRISMO	17 (34)	33 (66)		50 (100)		
All	157 (50)	472 (51.8)		629 (51.3)		
Age, mean (SD)						
Army STARRS	25.8(5.1)	25.5(5.2)	7.54E-01	25.6(5.2)		
DNHS	51.6 (11.1)	55.6 (17.1)	7.66E-02	55.3 (16.8)		
GTP	41.7 (11.4)	42.4 (12.5)	5.48E-01	42.2 (12.1)		
MRS I&II	23.3 (2.3)	22.9(1.9)	3.59E-01	23.1(2.1)		
PRISMO	28.1 (10.1)	27.5(9.1)	8.29E-01	27.7(9.3)		
All	36.1 (13.4)	44.1 (18)	5.89E-16	42.1 (17.3)		
PTSD symptom severity, mean (SD)						
Army STARRS	56.9 (9.6)	22.4 (5.8)	4.83E-28	32 (17)		
DNHS	63 (16)	32.7 (11.4)	1.89E-11	34.9 (14.2)		
GTP	70.4 (18.6)	25.1(16.9)	2.17E-32	38.5 (27.1)		
MRS I&II	65.4 (14.8)	13.6(11.8)	1.30E-42	40.2 (29.2)		
PRISMO	42 (4.4)	27(4.8)	6.72E-13	32.1(8.5)		
All	63.1(16.7)	27.7 (13.3)	5.88E-88	35.8 (20.5)		
Self-reported Race/Ethnicity, N (%)						
Army STARRS						
African American	3(2)	12(7.8)		15(9.8)		
White	29 (19)	88 (57.5)		117(76.5)		
Other	10(6.5)	11(7.2)		21 (13.7)		
DNHS						
African American	28 (6.7)	381 (91.6)		409 (98.3)		
Other	3(0.7)	4(1)		7(1.7)		
GTP						
African American	153 (31.6)	307 (63.4)		460 (95)		
Other	8(1.7)	16(3.3)		24(5)		
MRS I&II						
African American	2(1.6)	2(1.6)		4(3.3)		
White	53 (43.1)	53 (43.1)		106 (86.2)		
Other	8(6.5)	5(4.1)		13 (10.6)		
PRISMO						
African American	1(2)	1(2)		2(4)		
White	11(22)	27 (54)		38 (76)		
Other	5(10)	5(10)		10(20)		

Table 1 Demographic and clinical characteristics of the studies included in the discovery cohort

Table 1 (continued)

minf R package [\[20](#page-12-17)] to eliminate any sex-discordant samples. Quality control (QC) was performed on each cohort separately, using a standardized pipeline as previously described [\[21](#page-12-18)]. A total of 818,691 probes passed QC. Normalization was carried out using the single-sample Noob (ssNoob) method in the *minf* R package [\[20](#page-12-17)]. Furthermore, *ComBat* adjustment was performed, using an empirical Bayesian framework implemented in the *SVA* R package [\[22](#page-12-19), [23\]](#page-12-20) to reduce the likelihood of bias due to known batch efects (chip and position), while preserving the variation for age, sex (if applicable), and PTSD. The resulting QC'd data was used in subsequent analyses.

Estimation of covariates *Smoking scores*

Studies have linked methylation at many genomic loci to smoking status $[24-29]$ $[24-29]$ $[24-29]$. Therefore, to adjust for DNAm diferences related to smoking, we calculated smoking scores from DNAm data based on the weights obtained from 39 CpGs located at 27 loci, as previously described [[30\]](#page-12-23).

Cell proportions

It is important to consider cellular heterogeneity in epigenome-wide association studies (EWAS) [[31](#page-12-24)] since whole blood contains various cell types, each with its own DNAm profle [\[32,](#page-12-25) [33\]](#page-12-26). To address this, cell proportions (CD4+T, CD8+T, Natural Killer (NK), B-cells, monocytes, and neutrophils) were estimated using reference data [\[34](#page-13-0)] and the Robust Partial Correlation (RPC) method implemented in the *EpiDISH* R package [\[35\]](#page-13-1).

Ancestry principal components

Several studies have found variations in DNAm levels among diferent populations (race/ethnicity) at certain CpG sites $[36-41]$ $[36-41]$. Therefore, to account for population stratifcation, ancestry principal components (PCs) were generated from methylation data using a subset of CpGs in close proximity to SNPs in data from the 1000 Genomes Project [\[42,](#page-13-4) [43](#page-13-5)]. As previously reported [\[42](#page-13-4), [43\]](#page-13-5), PC 2 and 3 were the components most correlated with ancestry and thus, used to adjust for population stratifcation in this study.

Covariate adjustment

All the discovery cohorts had a small percentage of missing values (ranging from 0.002 to 0.03%). As the machine learning models require complete data, we used the mean method—a common and simple imputation technique to impute the missing data while maintaining the distribution of the data [\[44](#page-13-6), [45](#page-13-7)]. We then adjusted the DNAm data for potential confounding factors, including cell composition, ancestry, smoking score, sex (if applicable), and age, for models 1 and 2 (described below). The adjustment was made for each CpG by regressing out all the covariates using linear regression and then replacing the values of CpG with the corresponding residuals [\[46](#page-13-8)]. For model 3 (described below), we also regressed out the two exposure variables of interest, cumulative trauma and childhood trauma, in addition to the covariates used in models 1 and 2. This was done separately for each cohort to account for any diferences related to exposure variables in individual cohorts.

Analysis

Overall approach

Our goal was to develop a series of models based on important (i.e. set of features with best classifcation accuracy) methylation- and (in some cases) exposurerelated features to classify PTSD that would then be used to derive risk scores with which to predict PTSD. To train the models, we utilized unique, trauma exposed participants from the discovery cohort in a cross-sectional approach. Model 1 was designed to classify PTSD by including two exposure variables—cumulative trauma (number of traumatic events experienced) and childhood trauma (experienced at<18 years of age)—along with DNAm data, as increasing levels of exposures are known to substantially increase the risk of developing PTSD [\[47](#page-13-9)[–50](#page-13-10)] and were thus hypothesized to contribute high predictive power to our model. The purpose of Model 2 was to classify PTSD using only DNAm data, without relying on the discriminatory power of cumulative trauma or childhood trauma; this model would enable potential application to cohorts in which only DNAm data were available. Model 3 was developed with a unique purpose, distinct from Model 1. Namely, it was created to account for variations in exposure variables among individual cohorts. In this model, exposure variables were intentionally excluded from the analysis because they were used as covariates in DNAm data adjustment. While Model 3 addresses the challenge of cohort-specifc variations, it does not possess the same predictive power as Model 1, which incorporates these exposure variables. The adjusted data was then subjected to the following analysis processes.

Feature selection and scaling

We used *SelectKBest* in *Scikit-learn* [\[51\]](#page-13-11), a univariate feature selection approach. This method computes ANOVA F-values based on univariate statistical tests to identify the best features in relation to a particular phenotype*.* We identifed the most important features from DNAm and exposure variables (in cases of Models 1 & 2) based on the rank order of the features' association with PTSD. For Model 3, we selected features solely from DNAm data. The feature selection process was repeated 500 times, ranging from 10 to 5000 features with a 10-feature increment each time to determine the optimal feature set for the Elastic Net model best accuracy. As diferent studies/ cohorts used diferent instruments to measure cumulative trauma and childhood trauma, we normalized the data using a min–max scale that ranged from [[1](#page-12-0)].

Training and testing

In order to identify the best model to classify PTSD and determine risk scores, we trained three popular machine learning models —Random Forest, Lasso, and Elastic Net on 75% of the data, and then tested them on the remaining 25% using the *Scikit-learn* [\[51](#page-13-11)] framework. We also conducted a tenfold cross-validation on training and testing data to evaluate the efectiveness of the models (Figure S1.1). After selecting the most accurate machine learning model, which was evaluated based on the methylation and exposure variable dataset, we used important features identifed during the feature selection process to classify PTSD. Following covariate adjustment of the two additional exposure variables, we re-ran the

feature selection process to identify important features for Model 3 (described below). Performance of the models was assessed using accuracy, precision, recall, f1-score and area under the curve (AUC) metrics.

Risk scores

Risk scores are the weighted sum of the important features. Using feature weights (i.e. efect sizes) from training data (75%), we created risk scores using discovery cohort test data (25%), in order to test for an association between risk scores and PTSD. Model 1 contributed to the development of exposure and methylation risk scores (eMRS), whereas Model 2 provided methylation-only risk scores (MoRS). Finally, Model 3 led to the creation of methylation-only risk scores with adjusted exposure variables (MoRSAE).

A logistic model was employed to test for an association between risk scores (eMRS, MoRS and MoRSAE) and PTSD, and the Nagelkerke approach was used to assess the models' resulting R-Squared (R^2) values. For all analyses, a Wilcoxon rank-sum test was used to assess diferences in risk scores between cases and controls. To assess the direction of efect and strength of association among study variables in the both discovery and independent cohorts, Pearson's and point-biserial correlation was used, as appropriate.

Independent validation

To validate the risk scores, we tested their ability to distinguish those with vs. without PTSD in four independent, external cohorts using the same pre-processing and covariate adjustment pipeline as in the discovery cohort. Brief descriptions of the external cohorts (NCPTSD-TRACTS, BEAR, DCHS and PROGrESS) are provided in Supplementary File 1. We utilized weights from significant features identifed in models 1, 2, and 3 of the discovery cohort to generate risk scores (i.e., eMRS, MoRS, and MoRSAE) in the external cohorts. Similar to the discovery cohorts, we conducted Pearson and Point-Biserial correlation tests, association tests using logistic regression model, and Wilcoxon rank-sum tests on external cohorts.

MRS-based predictive analyses

In cohorts with available pre-deployment data, a logistic model was used to predict post-deployment PTSD using risk scores calculated from *pre-deployment* DNAm data and exposure data (Army STARRS, MRS Iⅈ $n=276$). Note that these participants had their post-deployment DNAm data included in the discovery cohort analyses described above.

Enrichment analysis

To investigate the biological signifcance of the important CpGs identifed in the feature selection step, we performed Gene Ontology (GO) enrichment analysis and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway analysis using *missMethyl* [\[52](#page-13-12)]. Gene ontologies and KEGG pathways that reached a nominal signifcance level of *p*<0.05 were considered important.

Results

Description of discovery cohort

Table [1](#page-2-0) provides a summary of the demographic characteristics and clinical information of all participants (n=1226) in the discovery cohort with current PTSD. More information about cumulative and childhood trauma is provided in Table S1. A slight majority of participants were male $(n=629)$. Two cohorts, DNHS and GTP, were comprised mostly of African Americans, while the remaining three cohorts were predominantly of European ancestry. In all cohorts, a signifcant diference in PTSD symptom severity was observed between cases and controls $(p < 0.05)$. With the exception of Army STARRS, childhood trauma also demonstrated a signifcant diference between PTSD cases and controls $(p<0.05)$ in all cohorts. Finally, a signifcant diference was observed in cumulative trauma between cases and controls in DNHS and GTP ($p < 0.001$).

Development of methylation risk scores to distinguish those with vs. without PTSD

We developed three diferent risk scores with the goal of distinguishing those with vs. without PTSD using machine learning approaches. Our frst model, eMRS, included both exposure and DNAm variables and identifed 3730 features (3728 CpGs, cumulative trauma, and childhood trauma) as important in the discovery cohort. Using these 3730 features, Elastic Net approaches were employed to achieve the best accuracy (92%; Fig. [1](#page-5-0)), pre-cision (91%), recall (87%), and f1-score (89%); Table [2](#page-5-1) (See Fig. S2 for AUCs with Lasso and Random Forest approaches). The eMRS significantly predicted PTSD (beta=2.64, $p < 0.001$), $R^2 = 0$ 0.70), with higher eMRS values in PTSD cases than controls $(p<0.001;$ Fig. [2](#page-6-0)A, left plot). Our second MoRS model, based solely on the 3728 methylation features in model 1, accurately classifed PTSD with 89% accuracy and had an AUC of 95% (Fig. [3;](#page-6-1) Table [2](#page-5-1)). Additionally, the precision, recall, and f1-score were at 86%, 83%, and 84%, respectively, as shown in Table [2](#page-5-1). As with eMRS, the MoRS significantly predicted PTSD (beta=2, $p < 0.001$, $R^2 = 0.54$) and had higher MoRS values in cases vs controls $(p < 0.001)$ (Fig. [2A](#page-6-0), middle plot). Our third and fnal model (i.e.,

Fig. 1 The confusion matrix for Model 1 displays an accuracy of 92% on test data (N=307), while the ROC curve indicates an AUC of 96% during the tenfold cross-validation using all data ($N=1226$)

Table 2 Elastic net model performance across the three models

Model	Accuracy (%)	Precision (%)	Recall (%)	F1-score (%)	AUC (%)
eMRS (Model 1)	92	9.	87	89	96
MoRS (Model 2)	89	86	83	84	95
MoRSAE (Model 3)	84	80			89

MoRSAE), which used DNAm data adjusted for the two exposure variables as well as the other covariates in models 1 and 2, identifed 4150 signifcant features that classifed PTSD with 84% accuracy and an AUC of 89% (Fig. [4](#page-7-0), with precision, recall, and f1-score at 80%, 77%, and 78%, respectively (Table [2](#page-5-1)). As with models eMRS and MoRS, MoRSAE signifcantly predicted PTSD (beta=1.20, $p < 0.001$, $R^2 = 0.36$) and had significantly ($p < 0.001$) different, and higher, MoRSAE in PTSD cases vs. controls (Fig. [2A](#page-6-0), right plot). In summary, while all three models produced risk scores that signifcantly predicted PTSD in the test dataset, and showed higher scores in aggregate between cases and controls, there was a decline in effect size (b) and explanatory power (R^2) such that eMRS>MoRS>MoRSAE.

Intercorrelation among study variables

A signifcant positive point-biserial correlation between eMRS and current PTSD was observed $(p=0.72)$, $p < 0.001$; Figure S3). Cumulative trauma ($p = 0.40$, *p*<0.001) and childhood trauma (ρ =0.57, *p*<0.001) also showed a positive and signifcant correlation with eMRS. Notably, there was also a signifcant and positive point-biserial correlation (ρ=0.62, *p*<0.001) between MoRS and PTSD, signifcant and positive correlation between cumulative trauma and MoRS (ρ = 0.16, p < 0.01) and childhood trauma and MoRS (ρ =0.169, p <0.01) (Figure S3). In contrast, while we observed a signifcant $(p<0.001)$ and positive point-biserial correlation $(p=0.49)$ between MoRSAE and PTSD (Figure S3), we observed a negative correlation between MoRSAE and cumulative trauma ($p = -0.13$, $p = 0.02$) and childhood trauma (ρ = -0.12, p = 0.03), respectively.

Validation of risk scores in external cohorts

We conducted external validation on risk scores from the three diferent models across four external cohorts— NCPTSD-TRACTS, BEAR, DCHS and PROGrESS. The NCPTSD-TRACTS cohort demonstrated a noticeable distinction ($p < 0.05$) in childhood trauma, but not in cumulative trauma (Table S1) between cases and controls. Similar to the discovery cohorts, the BEAR cohort exhibited a signifcant diference in both cumulative trauma and childhood trauma when comparing cases and controls. The DCHS cohort, on the other hand, only showed a signifcant diference in cumulative trauma, while the PROGrESS cohort did not display any

Fig. 2 Distribution and variation of risk scores between cases and controls in test data (N=307) in fgure legend, 0 is No PTSD and 1 is PTSD. **A**) The distribution of risk scores for Models 1, 2, and 3 is shown for both cases and controls. **B**) The diference in risk scores, and associated p value, between cases and controls is displayed. Model 1 calculates exposure and methylation risk scores (eMRS), while Model 2 calculates risk scores based only on methylation variables (MoRS). Model 3 calculates risk scores based on methylation variables adjusted for exposure variables (MoRSAE). The risk scores are higher in PTSD cases compared to controls. The Wilcoxon test confrms a signifcant diference in risk scores between cases and controls with *p*<0.001 for all models (1, 2, and 3)

Fig. 3 The confusion matrix for Model 2 displays an accuracy of 89% on test data (N=307), while the ROC curve indicates an AUC of 95% during the tenfold cross-validation using all data (N=1226)

signifcant diference in trauma variables between cases and controls.

The eMRS significantly predicted PTSD in one external cohort, BEAR (beta=0.6839, *p*=0.006) (Table S2); in this cohort, there was also a signifcant correlation (ρ =0.24, p =0.003) between eMRS and PTSD (Figure S4) and a signifcant diference in eMRS between PTSD cases and controls $(p=0.02,$ Figure S5). The eMRS did

Fig. 4 The confusion matrix for Model 3 displays an accuracy of 84% on test data (N=307), while the ROC curve exhibits an AUC of 89% during the tenfold cross-validation process using all data (N=1226)

not signifcantly predict PTSD in any of the other three independent cohorts; however, the correlation between eMRS and PTSD showed the same (i.e., positive) direction of efect in the NCPTSD-TRACTS (beta=0.0598, $p = 0.35$), PROGrESS (beta=0.1141, $p = 0.53$) and DCHS (beta = 0.0631 , $p = 0.81$) cohorts (Figures S6-S11). For model 2, the MoRS did not signifcantly predict PTSD in any external cohort (NCPTSD-TRACTS: beta=-0.0977, *p*=0.28; BEAR: beta=0.0239, *p*=0.93; PROGrESS: beta=0.2156, *p*=0.52; DCHS: beta=0.3739, *p*=0.37). On the other hand, for model 3, the MoRSAE approached signifcance in association with PTSD in the NCPTSD-TRACTS cohort (beta = -0.1707 , $p = 0.05$) and had significant diference in risk scores between cases and controls $(p=0.018)$ (Figure S7); however, the direction of effect was opposite to that observed in the discovery cohort.

Testing of pre-deployment risk scores to predict future PTSD

A compelling feature of risk scores is their ability to predict future disease risk. In our data, we were able to test the predictive ability of the MRS derived from our diagnostic/classifcation models on prospective risk of PTSD in two of our pre-deployment military cohorts, MRS I&II and Army STARRS (with data from the two cohorts analyzed together). MRS were calculated using "unseen" DNAm data from a pre-deployment timepoint, i.e. using DNAm data not included in the discovery cohort. All three models signifcantly predicted future PTSD based on risk scores calculated with pre-deployment data (eMRS beta = 1.92, $p < 0.001$, $R^2 = 0.53$; MoRS beta = 1.99, $p < 0.001$, $R^2 = 0.46$; and MoRSAE beta=1.77, $p < 0.001$, R^2 =0.47) and had significant difference in risk scores between individuals who developed PTSD and those who did not (Figs. [5](#page-8-0), [6,](#page-9-0) [7](#page-10-0)).

Assessment of biological signifcance among Important CpGs

Gene ontology (GO) analysis on the set of 3728 CpGs from models 1 and 2 revealed 403 nominally signifcant GO terms; among the 4150 important CpGs from Model 3, 382 nominally signifcant GO terms were identified. There were 115 GO terms common between models, including regulation of muscle adaptation, positive regulation of autophagy of mitochondrion, and sucrose metabolic process. Additionally, the Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway analysis identifed 47 pathways for models 1 and 2 and 25 pathways for model 3 at *p*<0.05 (list of GO and KEGG terms are provided in Supplementary File 2). Further, 14 pathways were common in models 1 and 2, and model 3, including, HIF-1 signaling pathway, mTOR signaling pathway, Insulin signaling pathway and Galactose metabolism. None of the GO terms or KEGG pathways passed the multiple hypothesis correction test.

Discussion

It is crucial to identify individuals who are at a higher risk of developing PTSD in order to provide timely preventive measures and efective therapeutic interventions. MRS ofer dynamic and modifable genomic-based insights into disease risk. In this study, we leveraged machine learning and a diverse set of cohorts to develop MRS for PTSD, with an initial aim of distinguishing those with vs.

Fig. 5 Distribution and difference in risk scores (eMRS) between PTSD cases and controls pre- and post-deployment (N=262) — in figure legend, 0 is No PTSD and 1 is PTSD. **A**) The distribution of risk scores revealed that individuals who developed PTSD post-deployment had higher scores compared to those who did not, both before and after deployment. **B**) The diference in risk scores showed there was a signifcant (*p*<0.001) diference in risk scores in those with PTSD post-deployment using Wilcoxon test

without PTSD and, subsequently to predict future PTSD cases. MRS derived from three diferent models demonstrated both high precision and high accuracy in predicting PTSD (i.e., identifying probable PTSD cases vs. controls) in the test dataset and, moreover, signifcantly predicted future PTSD. Although our approach did not yield MRS that consistently predicted PTSD in independent cohorts, our work leverages data from a diverse set of cohorts to develop what is, to our knowledge, the frst methylation-based risk scores for PTSD. Future work that builds on this approach will help to advance personalized preventive strategies and therapeutic interventions for PTSD in order to reduce the impact of this debilitating disorder on individuals and society.

Among the three models tested, the eMRS model showed the highest accuracy and precision to classify PTSD by using both exposure and DNAm variables. The inclusion of exposure variables substantially adds to the predictive power of the model. This finding aligns with the literature that suggests that experiencing trauma, particularly during childhood, signifcantly increases the likelihood of developing PTSD [\[47,](#page-13-9) [53](#page-13-13), [54\]](#page-13-14). It is noteworthy that, despite not including any trauma exposure factors, the second model (MoRS) and third model

(MoRSAE) that solely utilized methylation data in training still displayed notable predictive ability in the test dataset. These findings suggest that, even without using trauma variables in prediction, DNAm can still provide significant predictive information about PTSD. This also emphasizes the signifcant impact that trauma can have on the epigenetic landscape, which is consistent with other research studies [[55,](#page-13-15) [56](#page-13-16)] that reported methylation diferences linked to trauma. Overall, the decrease in classifcation accuracy across the models in the test dataset, from eMRS to MoRSAE, highlights the crucial role and discriminatory power that both DNAm and trauma exposure have in classifying PTSD.

Our attempts to validate the three models showed variable results across models and cohorts. The eMRS signifcantly predicted PTSD in one cohort, BEAR, with the same direction of efect as in the discovery cohort; the MoRSAE approached signifcance in predicting PTSD (*p*=0.05) in the NCPTSD-TRACTS cohort for MoRSAE, although with an opposite direction of efect to the discovery cohort. This variability may be due to individual diferences in the type or severity of trauma in each cohort. For example, similar to the discovery sample, the BEAR cohort showed signifcantly higher

Fig. 6 Distribution and difference in risk scores (MoRS) between cases and controls pre- and post-deployment (N=262) — in figure legend, 0 is No PTSD and 1 is PTSD. **A**) Distribution of risk scores between cases and controls. Risk scores are higher in those who developed PTSD post-deployment than who didn't in both pre and post deployment. **B**) Diference in risk scores between cases and controls. Wilcoxon test showed a signifcant difference ($p < 0.001$) in risk scores between cases and controls

levels of both cumulative and childhood trauma in participants with vs. without PTSD—a pattern not observed in any of the other three validation cohorts (Table S1). While we attempted to account for variability in trauma exposure by regressing out these efects in our MoR-SAE model, this did not improve the validation results. These results suggest that it may be necessary to develop a trauma-specifc MRS in order to more precisely capture the infuence of trauma, and its variability, in relation to classifcation and prediction of PTSD risk that generalizes across cohorts. We acknowledge that smaller sample sizes like the BEAR cohort can increase the risk of false positives. Still, the signifcant correlation and prediction results suggest that the observed strong association between eMRS and PTSD in the BEAR cohort is less likely due to chance alone. We emphasize the need for future studies with larger discovery and validation datasets to confrm our fndings and further explore the observed association.

The ability to predict PTSD prior to deployment is particularly important, as deployment is linked to a higher probability of trauma exposure than typically observed in community samples and higher trauma load increases risk for PTSD [\[54](#page-13-14)]. All three models signifcantly predicted future development of PTSD based on pre-deployment data, which is notable because these data preceded trauma exposure and were not included in the training or testing phase of MRS model development. This suggests that classifcation-based MRS may be useful in predicting risk for future PTSD in populations with anticipated trauma exposure.

Previous work has leveraged DNAm data as one among many biomarker types included in risk score approaches to predicting PTSD [\[57](#page-13-17), [58\]](#page-13-18). An earlier study focused on war zone-related PTSD identifed a set of 343 candidate biomarkers, of which 98 were DNAm values associated with particular genes [\[57](#page-13-17)]. From our identified list of signifcant CpGs (3728 in models 1 and 2), cg16335858 in *GYLTL1B* (*Glycosyltransferase-like 1B*) was previously identifed as a biomarker in diagnosing war zone-related PTSD [\[57\]](#page-13-17). From the list of 4150 CpGs (model 3), one additional CpG, cg25448062 in *FADS1* (*Fatty acid desaturase 1)* was identifed as a diagnostic biomarker in the same study. A subsequent study [\[58](#page-13-18)] showed prediction

Fig. 7 Distribution and difference in risk scores (MoRSAE) between cases and controls pre- and post-deployment (N=262) — in figure legend, 0 is No PTSD and 1 is PTSD. **A**) The distribution of MoRSAE is higher in those who developed PTSD post-deployment **B**) The diference in risk scores showed there was a signifcant (*p*<0.001) diference in risk scores in those with PTSD post-deployment using Wilcoxon test

of post-deployment PTSD symptoms with the best AUC of 88% and CpGs cg01208318 and cg17137457 as top predictors but none of these were replicated in our study. More broadly, it is interesting to note that, 4 CpGs (cg04583842, cg04987734, cg16758086 and cg19719391) in genes *BANP*, *CDC42BPB*, *CHD5* and *Intergenic* respectively, have been associated with PTSD in recent PGC EWAS meta-analyses (Katrinli et al., submitted). Our results build on these earlier studies, highlighting novel CpGs that, when combined in a weighted, risk score format, may contribute to PTSD prediction.

In this study, there were no results from GO or KEGG pathway analyses that remained signifcant following multiple hypothesis testing; however, the GO terms and KEGG pathways shared among the three models provide interesting clues about the biological mechanisms that may be involved in the development of PTSD. For example, positive regulation of autophagy of mitochondrion, identifed as nominally signifcant biological processes in all three models, is noteworthy, as prior research has suggested that autophagy plays a role in neurodegenerative illnesses [[59–](#page-13-19)[61\]](#page-13-20), and exploring its connection to PTSD could provide insights into the disorder's neurobiological

underpinnings. Additionally, the link to sucrose metabolic process is intriguing and raises questions about the relationship between energy metabolism and stress responses [\[61](#page-13-20)], as metabolic disorders have been associated with PTSD [\[63](#page-13-21)]. KEGG pathway analyses revealed additional implicated pathways, including mTOR and insulin signaling, which play a crucial role in cellular growth and metabolism, highlighting the extensive physiological efects of PTSD beyond psychological distress [[63,](#page-13-21) [64](#page-13-22)].

Our study is not without limitations. Chief among these is our external validation results, which showed validation for only one model in one of the four cohorts tested. To date, attempts to validate risk scores in external, independent cohorts–as done in this study–are not common, and most work focusses on reporting results based on validation in a test (i.e., internal) dataset [[14](#page-12-11)]. Results from this work highlight the need to increase efforts to do so, in order to arrive at robust, generalizable MRS with the potential for future clinical application. While our three classifcation-based MRS models showed good prediction of future PTSD in pre-deployment data, it is unclear whether they would perform

as well in predicting future PTSD in civilian populations. As more data become available, the inclusion of additional molecular, environmental, and psychosocial factors in MRS scores may enhance their accuracy in predicting the condition and, relatedly, improve their performance in independent cohorts.

Supplementary Information

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Supplementary Material 1. Supplementary Material 2.

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Availability of data and materials

Owing to military cohort data sharing restrictions, data from MRS I& II, Army STARRS, PRISMO, and NCPTSD-TRACTS cannot be publicly posted. For other cohorts, individual-level data from the cohorts or cohort-level summary statistics may be made available to researchers following an approved analysis proposal through the PGC Post-traumatic Stress Disorder EWAS group with agreement of the cohort PIs. For additional information on access to these data, including PI contact information for the contributing cohorts, please contact the corresponding author.

Declarations

Ethics approval and consent to participate

All participants included in analyses of the discovery and external validation cohorts provided informed consent for their participation. Details on the

specific IRB approval associated with each study are provided in the Supplementary Material.

Consent for publication

Not applicable.

Competing interests

Murray B. Stein has in the past 3 years received consulting income from Acadia Pharmaceuticals, Aptinyx, atai Life Sciences, BigHealth, Biogen, Bionomics, BioXcel Therapeutics, Boehringer Ingelheim, Clexio, Delix Therapeutics, Eisai, EmpowerPharm, Engrail Therapeutics, Janssen, Jazz Pharmaceuticals, Neu‑ roTrauma Sciences, PureTech Health, Sage Therapeutics, Sumitomo Pharma, and Roche/Genentech. Dr. Stein has stock options in Oxeia Biopharmaceuticals and EpiVario. He has been paid for his editorial work on Depression and Anxiety (Editor-in-Chief), Biological Psychiatry (Deputy Editor), and UpToDate (Co-Editor-in-Chief for Psychiatry). He has also received research support from NIH, Department of Veterans Afairs, and the Department of Defense. He is on the scientifc advisory board for the Brain and Behavior Research Foundation and the Anxiety and Depression Association of America. Dr. Chia-Yen Chen is an employee of Biogen. Dr. Nikolaos P. Daskalakis has served on scientifc advisory boards for BioVie Pharma, Circular Genomics and Sentio Solutions for unrelated work. Dr. Nicole R. Nugent is a member of the scientifc advisory board for Ilumivu. Dr. Sheila Rauch support from Wounded Warrior Project (WWP), Department of Veterans Afairs (VA), National Institute of Health (NIH), McCormick Foundation, Tonix Pharmaceuticals, Woodruf Foundation, and Department of Defense (DOD). Dr. Rauch also receives royalties from Oxford University Press and American Psychological Association Press. Dr Ressler reported receiving personal consulting fees from Sage Therapeutics, Senseye, Boerhinger Ingelheim, Jazz Pharmaceuticals, and Acer, Inc. and a sponsored research grant from Alto Neuroscience outside the submitted work.

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